

# NeoBio Mount Fluo with DAPI

(Fluorescent Mounting Medium with 4,6-diamidino-2-phenylindole)

#Cat: NB-23-00159-1 #Cat: NB-23-00159-2 Size: 100ml liquid (ready-to-use) Size: 18ml liquid (ready-to-use)

## Description

Fluorescent mounting medium with DAPI is an aqueous mounting medium for preserving fluorescence of tissue and cell smears. This unique formula prevents rapid photobleaching of FITC, Texas Red, AMCA, Cy2, Cy3, Cy5, Alexa Fluor<sup>™</sup> 488, Alexa Fluor<sup>™</sup> 594, Green fluorescent protein (GFP), Tetramethylrhodamine (TRITC), Redox. Phycoerythrin (RP-E), Phyocyanin (PC), and Allophycocyanin (APC). Fluorescence is retained during prolonged storage at 4°C in the dark. This medium does not contain phenylenediamine, which destroys immunofluorescence of Cy dyes, RP-E, PC and APC.

This mounting medium is fortified with DAPI which is a counter-stain for DNA. This product is to be used in in situ hybridization techniques or other methods where fluorescence of DNA staining is required. DAPI excites at 360nm and emits at 460nm, producing a blue fluorescence. RNA is also stained with DAPI.

### Intended Use

Immunofluorescence, confocal microscopy.

## Storage

Store at 2-8°C. Protect from light, DO NOT FREEZE.

### **Recommended Protocol**

- 1. Bring the vial to room temperature.
- 2. Rinse slide with DISTILLED OR DEIONIZED WATER, touch the edges of slide on a paper towel to remove excess water. Place slides on a flat surface away from light.
- 3. Turn the vial upside down and open the dropper to remove any air bubbles.
- 4. Apply 3-4 drops of mounting medium directly on top of the specimen and spread out evenly by tilting back and forth or spread evenly with a 0.2 ml plastic pipette tip making sure the tissue is not touched. Excess medium can be removed by touching the edges of the slide on a paper towel.
- 5. Let stand at room temperature for about 5 minutes.
- 6. Apply coverslip, carefully avoiding air bubbles.
- 7. The specimen is ready for visualization under a microscope.
- 8. One can seal the edges of the cover slip with nail polish, any organic mounting medium or our O-mount. If a coverslip is not used, air bubbles will appear in few days.
- 9. The fluorescent mounting medium helps to preserve fluorescence for longer peroid if the slide is stored in the dark at 2-8°C.



#### Removal of Coverslip:

Coverslip can be removed before sealing the edges. Soak slide in warm (37°C) water for a few minutes. Carefully and slowly move the coverslip. Soak in water for an additional few minutes to remove coverslip. Rinse slide several times with warm water to remove all mounting medium. The slide can be remounted again. additional few minutes to remove coverslip. Rinse slide several times with warm water to remove coverslip. Rinse slide several times additional few minutes to remove coverslip. Rinse slide several times with warm water to remove coverslip. Rinse slide several times with warm water to remove all mounting medium. The slide can be remounted again.

#### Remarks

Follow good laboratory practice to handle the procedure.

For Research Use Only. Not for Diagnostic or Therapeutic Use.