Product Manual

OxiSelect™ UV-Induced DNA Damage ELISA Kit (CPD Quantitation)

Catalog Number

STA-322 96 assays

STA-322-5 5 x 96 assays

FOR RESEARCH USE ONLY Not for use in diagnostic procedures



Introduction

Absorption of ultraviolet (UV) light produces two predominant types of DNA damage, cyclobutane pyrimidine dimers (CPD) and pyrimidine (6-4) pyrimidone photoproducts (6-4PP) (Figure 1). The result is a transition of C to T and CC to TT, which are the most frequent mutations of p53 in both human and mouse skin cancers. UV damaged DNA is usually repaired by nucleotide excision repair (NER) or base excision repair (BER). After UV exposure, cells activate p53 and stall the cell cycle for repair. If the damage is too severe, the cell will trigger apoptosis to get rid of DNA damaged, potentially mutant cells.

Cell Biolabs' OxiSelectTM Oxidative UV-induced DNA Damage ELISA Kit (CPD Quantitation) is an enzyme immunoassay developed for rapid detection and quantitation of CPD in any DNA samples. The quantity of CPD in unknown sample is determined by comparing its absorbance with that of a known CPD-DNA standard curve. Each kit provides sufficient reagents to perform up to 96 assays, including standard curve and unknown samples.

Figure 1: Structures of DNA lesions induced by UV Light

Assay Principle

CDP-DNA standards or unknown DNA samples are first heat denatured before being adsorbed onto a 96-well DNA high-binding plate. The CPDs present in the sample or standard are probed with an anti-CPD antibody, followed by an HRP conjugated secondary antibody. The CPD content in an unknown sample is determined by comparing with a standard curve that is prepared from predetermined CPD-DNA standards.

Related Products

- 1. STA-320: OxiSelectTM Oxidative DNA Damage ELISA Kit (8-OHdG Quantitation)
- 2. STA-321: OxiSelectTM DNA Double-Strand Break (DSB) Staining Kit
- 3. STA-323: OxiSelectTM UV-induced DNA Damage ELISA Kit (6-4PP Quantitation)
- 4. STA-324: OxiSelectTM Oxidative DNA Damage Quantitation Kit (AP sites)
- 5. STA-325: OxiSelectTM Oxidative RNA Damage ELISA Kit (8-OHG Quantitation)

Kit Components

1. DNA High-Binding Plate (Part No. 232404): One 96-well strip plate.



- 2. <u>DNA Binding Solution</u> (Part No. 232405): One 6 mL bottle.
- 3. Anti-CPD Antibody (Part No. 232202): One 20 µL vial.
- 4. Secondary Antibody, HRP Conjugate (Part No. 10902): One 50 μL vial.
- 5. Assay Diluent (Part No. 310804): One 50 mL bottle.
- 6. 10X Wash Buffer (Part No. 310806): One 100 mL bottle.
- 7. Substrate Solution (Part No. 310807): One 12 mL amber bottle.
- 8. Stop Solution (Part. No. 310808): One 12 mL bottle.
- 9. CPD-DNA Standard (Part No. 232203): One 100 μ L vial of 25 μ g/mL CPD-DNA in 1X TE Buffer.
- 10. <u>Reduced DNA Standard</u> (Part No. 232207): One 100 μL vial of 0.2 mg/mL reduced DNA in TE Buffer.

Materials Not Supplied

- 1. DNA samples such as cell or tissue genomic DNA
- 2. DNA Extraction Kit
- 3. Heating Block
- 4. PBS
- 5. 1X TE Buffer (10 mM Tris, pH 8.0, 1 mM EDTA)

Storage

Upon receipt, aliquot and store the Reduced DNA and CPD-DNA Standards at -20°C to avoid multiple freeze/thaw cycles. Store all other components at 4°C.

Preparation of Reagents

- 1X Wash Buffer: Dilute the 10X Wash Buffer Concentrate to 1X with deionized water. Stir to homogeneity.
- Anti-CPD Antibody and Secondary Antibody: Immediately before use dilute the Anti-CPD Antibody 1:1000 and Secondary Antibody 1:1000 with Assay Diluent. Do not store diluted solutions.

Preparation of Standard Curve

1. Convert CPD-DNA standard (25 μ g/mL) and Reduced DNA (200 μ g/mL) to single-stranded DNA by incubating the DNA at 95°C for 10 minutes and rapidly chilling on ice for 10 minutes.

Note: Aliquot and store denatured DNA at -20°C. Repeat the above denaturation step every time you prepare the CPD-DNA standard.

- 2. Freshly prepare 4 μg/mL of CPD-DNA by diluting the denatured 25 μg/mL stock in cold TE Buffer. Example: Add 8 μL to 42 μL of cold TE Buffer.
- 3. Freshly prepare 4 μ g/mL of Reduced DNA by diluting the denatured 200 μ g/mL stock in cold TE Buffer. Example: Add 40 μ L to 1.96 mL of cold TE Buffer.
- 4. Prepare a series of CPD-DNA standards according to Table 1.



Standard Tubes	4 μg/mL Denatured CPD- DNA (μL)	4 μg/mL Denatured Reduced DNA (μL)	CPD-DNA Conc. (ng/mL)
1	10	390	100
2	200 of tube #1	200	50
3	200 of tube #2	200	25
4	200 of tube #3	200	12.5
5	200 of tube #4	200	6.25
6	200 of tube #5	200	3.13
7	200 of tube #6	200	1.56
8	0	200	0

Table 1. Preparation of CPD-DNA Standards

Assay Protocol

- 1. Extract DNA from cell or tissue samples using a commercial DNA Extraction kit or other desired method.
- 2. Convert DNA sample to single-stranded DNA by incubating the sample at 95°C for 10 minutes and rapidly chilling on ice for 10 minutes.
- 3. Dilute DNA samples to $4 \mu g/mL$ in cold TE Buffer.

Note: Samples with high concentrations of CPD may be further diluted 2-4-fold in 4 μ g/mL Reduced DNA. A titration may be performed to ensure the samples fall in the range of the standard curve.

- 4. Add 50 μ L of unknown DNA samples or CPD-DNA standards to the wells of the DNA High-Binding plate.
- 5. Add 50 µL of DNA Binding Solution to each well. Mix well by pipetting and incubate at room temperature overnight on an orbital shaker. Each DNA sample including unknown and standard should be assayed in duplicate.
- 6. Remove the DNA solutions and wash twice with PBS. Blot plate on paper towels to remove excess fluid. Add 200 µL of Assay Diluent to each well and block for 1 hour at room temperature.
- 7. Remove the Assay Diluent. Blot plate on paper towels to remove excess fluid.
- 8. Add 100 μL of the diluted Anti-CPD Antibody to all wells and incubate for 1 hour at room temperature on an orbital shaker.
- 9. Wash 5 times with 250 μL of 1X Wash Buffer with thorough aspiration between each wash. After the last wash, empty wells and tap microwell strips on absorbent pad or paper towel to remove excess 1X Wash Buffer.
- 10. Add 100 μL of the diluted Secondary Antibody-HRP Conjugate to all wells and incubate for 1 hour at room temperature on an orbital shaker. Wash the strip wells 5 times according to step 9 above.
- 11. Warm Substrate Solution to room temperature. Add $100~\mu L$ of Substrate Solution to each well, including the blank wells. Incubate at room temperature on an orbital shaker. Actual incubation time may vary from 2-30 minutes.

Note: Watch plate carefully; if color changes rapidly, the reaction may need to be stopped sooner to prevent saturation.

- 12. Stop the enzyme reaction by adding 100 μL of Stop Solution to each well. Results should be read immediately (color will fade over time).
- 13. Read absorbance of each well on a microplate reader using 450 nm as the primary wave length. Use the Reduced DNA Standard as an absorbance blank.



Example of Results

The following figures demonstrate typical Oxidative UV-induced DNA Damage ELISA (CPD Quantitation) results. One should use the data below for reference only. This data should not be used to interpret actual results.

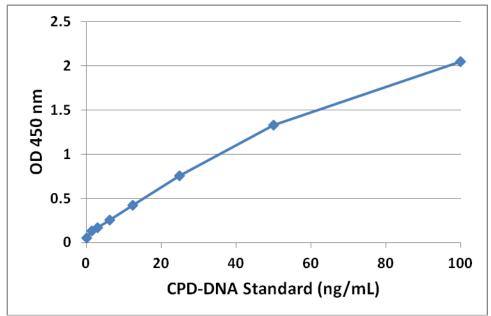


Figure 2: CPD-DNA Standard Curve.

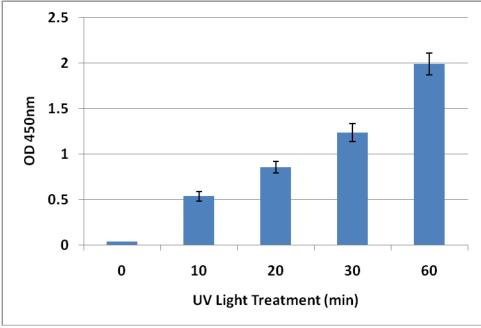


Figure 3: DNA Damage Induced by UV Light. 0.2 mg/mL Calf thymus DNA was exposed to UV light inside a cell culture hood for the time indicated. The CPD levels in denatured DNA samples were determined as described in the Assay Protocol.

References

1. Lippke JA, Gordon LK, Brash DE, Haseltine WA. (1981) *Proc Natl Acad Sci U S A.* **78**:3388–3392.



- 2. Mitchell DL, Nairn RS. (1989) Photochem Photobiol. 49:805–819.
- 3. Ananthaswamy HN, Loughlin SM, Cox P, Evans RL, Ullrich SE, Kripke ML. (1997) *Nat Med.* 3:510–514.
- 4. Soehnge H, Ouhtit A, Ananthaswamy ON. (1997) Front Biosci. 2: D538–D551.
- 5. el-Deiry WS, Tokino T, Velculescu VE, Levy DB, Parsons R, Trent JM, Lin D, Mercer WE, Kinzler KW, Vogelstein B. (1993) *Cell.* **75**:817–825.
- 6. Hermeking H, Lengauer C, Polyak K, He TC, Zhang L, Thiagalingam S, Kinzler KW, Vogelstein B. (1997) *Mol Cell.* 1:3–11.
- 7. Hill LL, Ouhtit A, Loughlin SM, Kripke ML, Ananthaswamy HN, Owen-Schaub LB. (1999) *Science*. **285**:898–900.

Recent Product Citations

- 1. Ivanova, I. et al. (2022). Improved Spectral Purity of 222-nm Irradiation Eliminates Detectable CPD Formation in Skin Reconstructs even at High and Repetitive Disinfecting Doses. *Photochem Photobiol*. doi: 10.1111/php.13594.
- 2. Polo, L.K. & Chow, F. (2022). Variation of antioxidant capacity and antiviral activity of the brown seaweed Sargassum filipendula (Fucales, Ochrophyta) under UV radiation treatments. *J. Appl. Phycol.* doi: 10.1080/26388081.2022.2030653.
- 3. Shin, J.Y. & Kang, N.G. (2022). Protective Effect of Glycyrrhiza glabra Extract on UV-induced Skin DNA Damage. *J. Soc. Cosmet. Sci. Korea.* doi: 10.15230/SCSK.2022.48.1.33.
- 4. Ciminera, A.K. et al. (2021). Elevated glucose increases genomic instability by inhibiting nucleotide excision repair. *Life Sci Alliance*. **4**(10):e202101159. doi: 10.26508/lsa.202101159.
- 5. Oliveira, B.R. et al. (2021). Inactivation of Aspergillus species in real water matrices using medium pressure mercury lamps. *J Photochem Photobiol B*. doi: 10.1016/j.jphotobiol.2021.112242.
- 6. Oliveira, B.R. et al. (2021). Light-emitting diodes effect on Aspergillus species in filtered surface water: DNA damage, proteome response and potential reactivation. *Environ. Pollut.* doi: 10.1016/j.envpol.2021.117553.
- 7. Wang, J. et al. (2021). Effects of TiO2-NPs pretreatment on UV-B stress tolerance in Arabidopsis thaliana. *Chemosphere*. **281**:130809. doi: 10.1016/j.chemosphere.2021.130809.
- 8. Xie, L. et al. (2020). Effects of artificial ultraviolet B radiation on the macrophyte Lemna minor: a conceptual study for toxicity pathway characterization. *Planta*. **252**(5):86. doi: 10.1007/s00425-020-03482-3.
- 9. Holzscheck, N. et al. (2020). Concomitant DNA methylation and transcriptome signatures define epidermal responses to acute solar UV radiation. *Sci Rep.* **10**(1):12918. doi: 10.1038/s41598-020-69683-8.
- 10. Shapira, Y. et al. (2019). Pre-germination plasma treatment of seeds does not alter cotyledon DNA structure, nor phenotype and phenology of tomato and pepper plants. *Biochem Biophys Res Commun*. doi: 10.1016/j.bbrc.2019.09.034.
- 11. Chazarin, B. et al. (2019). Limited Oxidative Stress Favors Resistance to Skeletal Muscle Atrophy in Hibernating Brown Bears (Ursus Arctos). *Antioxidants (Basel)*. **8**(9). pii: E334. doi: 10.3390/antiox8090334.
- 12. Innes, S.N. et al. (2019). Elevated air humidity increases UV mediated leaf and DNA damage in pea (Pisum sativum) due to reduced flavonoid content and antioxidant power. *Photochem Photobiol Sci.* **18**(2):387-399. doi: 10.1039/c8pp00401c.



- 13. Zhao, H. et al. (2018). Modulation of DNA Repair Systems in Blind Cavefish during Evolution in Constant Darkness. *Curr Biol.* **28**(20):3229-3243.e4. doi: 10.1016/j.cub.2018.08.039.
- 14. Cui, H. et al. (2016). The Stress-responsive Gene ATF3 mediates dichotomous UV responses by regulating the Tip60 and p53 proteins. *J Biol Chem.* **291**:10847-10857.
- 15. Rimann, M. et al. (2015). Standardized 3D bioprinting of soft tissue models with human primary cells. *J Lab Autom.* doi:10.1177/2211068214567146.
- 16. Donninger, H. et al. (2015). The RASSF1A tumor suppressor regulates XPA-mediated DNA repair. *Mol Cell Biol.* **35**:277-287.
- 17. Zirkin, S. et al. (2013). The PIM-2 Kinase is an essential component of the ultraviolet damage response that acts upstream to E2F-1 and ATM. *J. Biol. Chem.* **288**:21770-21789.
- 18. Burgess, H.M. et al. (2011). Nuclear relocalisation of cytoplasmic poly(A)-binding proteins PABP1 and PABP4 in response to UV irradiation reveals mRNA-dependent export of metazoan PABPs. *J. Cell Sci.* **124**: 3344-3355.

Warranty

These products are warranted to perform as described in their labeling and in Cell Biolabs literature when used in accordance with their instructions. THERE ARE NO WARRANTIES THAT EXTEND BEYOND THIS EXPRESSED WARRANTY AND CELL BIOLABS DISCLAIMS ANY IMPLIED WARRANTY OF MERCHANTABILITY OR WARRANTY OF FITNESS FOR PARTICULAR PURPOSE. CELL BIOLABS' sole obligation and purchaser's exclusive remedy for breach of this warranty shall be, at the option of CELL BIOLABS, to repair or replace the products. In no event shall CELL BIOLABS be liable for any proximate, incidental or consequential damages in connection with the products.

Contact Information

Cell Biolabs, Inc. 5628 Copley Drive San Diego, CA 92111

Worldwide: +1 858 271-6500 USA Toll-Free: 1-888-CBL-0505 E-mail: tech@cellbiolabs.com

www.cellbiolabs.com

©2011-2023: Cell Biolabs, Inc. - All rights reserved. No part of these works may be reproduced in any form without permissions in writing.

