## LSBiotm Mouse IgG ELISA Kit

Catalog No. LS-F10451

# **User Manual**

Please Read the Manual Carefully Before Starting your Experiment



For research use only. Not approved for use in humans or for clinical diagnosis.



www.lsbio.com

### Mouse IgG Antigen ELISA Kit

Strip well format. Reagents for up to 96 tests.

Rev: May 2014

#### **INTENDED USE**

This Mouse Immunoglobulin G (IgG) antigen assay is intended for the quantitative determination of total mouse IgG antigen in serum, plasma, hybridoma cell supernatants, ascites or other biological fluids. This assay does not distinguish IgG subclasses. For research use only.

#### **BACKGROUND**

IgG is the most abundant immunoglobulin in serum and is predominately involved in the secondary immune response. The IgG subclasses are designated 1, 2, 3 and 4 based on their relative prevalence in human serum.

#### **ASSAY PRINCIPLE**

Mouse IgG will bind to the affinity purified capture antibody coated on the microtiter plate. After appropriate washing steps, horseradish peroxidase labeled polyclonal anti-mouse IgG antibody binds to the captured protein. Excess antibody is washed away and TMB substrate is used for color development at 450nm. A standard calibration curve is prepared along with the samples to be measured using dilutions of mouse IgG. Color development is directly proportional to the concentration of total IgG in the samples.

#### **REAGENTS PROVIDED**

•96-well antibody coated microtiter strip plate (removable wells 8x12) containing anti-mouse IgG antibody, blocked and dried.

•10X Wash buffer: 1 bottle of 50ml

•5X Diluent: 1 bottle of 50ml

Mouse IgG standard: 1 vial lyophilized standard
 Anti-mouse horseradish peroxidase antibody: 1 vial

concentrated HRP labeled antibody
 TMB substrate solution: 1 bottle of 10ml

#### **STORAGE AND STABILITY**

Store all kit components at 4°C upon arrival. Return any unused microplate strips to the plate pouch with desiccant. Reconstituted standard may be stored at -80°C for later use. Do not freeze-thaw the standard more than once. Store all other unused kit components at 4°C. This kit should not be used beyond the expiration date.

#### **OTHER REAGENTS AND SUPPLIES REQUIRED**

- Microtiter plate shaker capable of 300 rpm uniform horizontally circular movement
- Manifold dispenser/aspirator or automated microplate washer
  - Microplate reader capable of measuring absorbance at 450 nm
- Pipettes and Pipette tips
- Deionized or distilled water
- •Polypropylene tubes for dilution of standard•Paper towels or laboratory wipes
- •1N H<sub>2</sub>SO<sub>4</sub> or 1N HCl

#### **PRECAUTIONS**

- FOR LABORATORY RESEARCH USE ONLY. NOT FOR DIAGNOSTIC USE.
- Do not mix any reagents or components of this kit with any reagents or components of any other kit. This kit is designed to work properly as provided.
- Always pour peroxidase substrate out of the bottle into a clean test tube. Do not pipette out of the bottle as contamination could result.
- Keep plate covered except when adding reagents, washing, or reading.
- •DO NOT pipette reagents by mouth and avoid contact of reagents and specimens with skin.
- •DO NOT smoke, drink, or eat in areas where specimens or reagents are being handled.

#### **PREPARATION OF REAGENTS**

- •1X Diluent: 5X Diluent may contain precipitate. Warm to redissolve before use. Dilute 50ml of 5X diluent concentrate with 200ml of deionized water.
- •1X Wash buffer: Dilute 50ml of 10X wash buffer concentrate with 450ml of deionized water.

#### **SAMPLE COLLECTION**

Collect plasma using EDTA or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000xg within 30 minutes of collection. Assay immediately or aliquot and store at  $\leq$  - 20°C. Avoid repeated freeze-thaw cycles.

#### **ASSAY PROCEDURE**

Perform assay at room temperature. Vigorously shake plate (300rpm) at each step of the assay.

#### **Preparation of Standard**

Reconstitute standard by adding 1ml of diluent directly to the vial and agitate gently to completely dissolve contents. This will result in a 1,000ng/ml standard solution.

Dilution table for preparation of mouse IgG standard:

production table for proparation of mouse igo standard.					
IgG	Dilutions				
concentration	Dilutions				
(ng/ml)					
500	500μl Diluent + 500μl (from std vial)				
250	500µl Diluent + 500µl (500ng/ml)				
100	600µl Diluent + 400µl (250ng/ml)				
50	500µl Diluent + 500µl (100ng/ml)				
25	500µl Diluent + 500µl (50ng/ml)				
10	600µl Diluent + 400µl (25ng/ml)				
5	500µl Diluent + 500µl (10ng/ml)				
2.5	500µl Diluent + 500µl (5ng/ml)				
1	600µl Diluent + 400µl (2.5ng/ml)				
0.5	500µl Diluent + 500µl (1ng/ml)				
0	500μl Diluent				
	Zero point to determine background				

NOTE: DILUTIONS FOR THE STANDARD CURVE AND ZERO STANDARD MUST BE MADE AND APPLIED TO THE PLATE IMMEDIATELY.

#### **Standard and Unknown Addition**

Remove microtiter plate from bag and add  $100\mu l$  IgG standards (in duplicate) and unknowns to wells. Carefully record position of standards and unknowns. Shake plate at 300rpm for 30 minutes. Wash wells three times with  $300\mu l$  wash buffer. Remove excess wash by gently tapping plate on paper towel or kimwipe.

NOTE: The assay measures IgG antigen in the 0.5-500 ng/ml range. If the unknown is thought to have high IgG levels, dilutions may be made in diluent. A

1:1,000,000 dilution for normal mouse plasma is suggested for best results.

#### **Antibody Addition**

Briefly centrifuge vial before opening. Dilute  $2\mu l$  of conjugated antibody in 10ml of diluent and add  $100\mu l$  to all wells. Shake plate at 300rpm for 30 minutes. Wash wells three times with  $300\mu l$  wash buffer. Remove excess wash by gently tapping plate on paper towel or kimwipe.

#### **Substrate Incubation**

Add 100 $\mu$ l TMB substrate to all wells and shake plate for 2-10 minutes. Substrate will change from colorless to different strengths of blue. Quench reaction by adding 50 $\mu$ l of 1N H<sub>2</sub>SO<sub>4</sub> or HCl stop solution to all wells when samples are visually in the same range as the standards. Add stop solution to wells in the same order as substrate upon which color will change from blue to yellow. Mix thoroughly by gently shaking the plate.

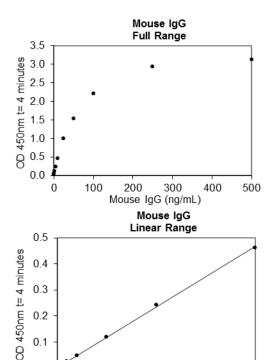
#### Measurement

Set the absorbance at 450nm in a microtiter plate spectrophotometer. Measure the absorbance in all wells at 450nm. Subtract zero point from all standards and unknowns to determine corrected absorbance (A<sub>450</sub>).

#### **Calculation of Results**

Plot A<sub>450</sub> against the amount of IgG in the standards. Fit a straight line through the linear points of the standard curve using a linear fit procedure if unknowns appear on the linear portion of the standard curve. Alternatively, create a standard curve by analyzing the data using a software program capable of generating a four parameter logistic (4PL) curve fit. The amount of IgG in the unknowns can be determined from this curve. If samples have been diluted, the calculated concentration must be multiplied by the dilution factor.

A typical standard curve (EXAMPLE ONLY):



#### **EXPECTED VALUES**

0.1 0.0 0

The concentration of IgG in normal mouse serum ranges from 5 to 12 mg/mL.

6

Mouse IgG (ng/mL)

10

#### PERFORMANCE CHARACTERISTICS

Sensitivity: The minimum detectable dose (MDD) was determined by adding two standard deviations to the mean optical density value of twenty zero standard replicates (range OD450: 0.05-0.059) and calculating the corresponding concentration. The MDD was 0.119 ng/ml. Intra-assay Precision: Three samples of known concentration were tested twenty times on one plate to assess intra-assay precision.

Sample	1	2	3
n	20	20	20
Mean (ng/ml)	1.84	15.9	141
Standard Deviation	0.116	0.700	12.8
CV (%)	6.29	4.41	9.10

**Inter-assay Precision:** Three samples of concentration were tested in ten independent assays to assess inter-assay precision.

Sample	1	2	3
n	10	10	10
Mean (ng/ml)	2.04	20.5	127
Standard Deviation	0.143	0.936	12.9
CV (%)	7.01	4.56	10.2

Recovery: The recovery of antigen spiked to levels throughout the range of the assay in blocking buffer was evaluated.

Sample	1	2	3	4	
n	4	4	4	4	
Mean (ng/ml)	1.97	7.44	32.4	147	
Average % Recovery	98	99	93	98	
Range	91- 109%	94- 102%	88- 100%	86- 109%	

Linearity: To assess the linearity of the assay, pooled mouse plasma samples containing high concentrations of antigen were serially diluted to produce samples with values within the dynamic range of the assay.

Sample	1:2	1:4	1:8	1:16	
n	4	4	4	4	
Average % of Expected	99	103	99	98	
Range	96- 104%	96- 107%	89- 109%	95- 101%	

Specificity: This assay recognizes total mouse IgG. Pooled normal plasma from rat, rabbit, sheep, pig, and human were assayed, and no significant cross reactivity was observed.

Sample Values: Samples were evaluated for the presence of the antigen at varying dilutions.

Sample Type	Dilution	Mean (mg/mL)		
	1:100,000	10.3		
	1:250,000	11.2		
Citrata Dlasma	1:500,000	10.5		
Citrate Plasma	1:1,000,000	8.68		
	1:2,000,000	10.9		
	1:4,000,000	8.89		

#### **DISCLAIMER**

This information is believed to be correct but does not claim to be all-inclusive and shall be used only as a guide. The supplier of this kit shall not be held liable for any damage resulting from handling of or contact with the above product.

#### **Example of ELISA Plate Layout**

96 Well Plate: 22 Standard wells, 74 Sample wells

	1	2	3	4	5	6	7	8	9	10	11	12
Α	0	0.5 ng/ml	1 ng/ml	2.5 ng/ml	5 ng/ml	10 ng/ml	25 ng/ml	50 ng/ml	100 ng/ml	250 ng/ml	500 ng/ml	
В	0	0.5 ng/ml	1 ng/ml	2.5 ng/ml	5 ng/ml	10 ng/ml	25 ng/ml	50 ng/ml	100 ng/ml	250 ng/ml	500 ng/ml	
С												
D												
ΕF												
G												
Н												

**Important Note:** During shipment, small volumes of product will occasionally become entrapped in the seal of the product vial. We recommend briefly centrifuging the vial to dislodge any liquid in the container's cap prior to opening.

**Warning:** This reagent may contain sodium azide and sulfuric acid. The chemical, physical, and toxicological properties of these materials have not been thoroughly investigated. Standard Laboratory Practices should be followed. Avoid skin and eye contact, inhalation, and ingestion. Sodium azide forms hydrazoic acid under acidic conditions and may react with lead or copper plumbing to form highly explosive metal azides. On disposal, flush with large volumes of water to prevent accumulation.

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